

## Cytokine Polymorphism and HLA Genotyping in Patients with Temporal Lobe Epilepsy Related to Hippocampal Sclerosis

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### ABSTRACT

**Objective:** Hippocampal sclerosis (HS) is the most common pathological substrate associated with mesial temporal lobe epilepsy (MTLE), where inflammatory processes are known to play an increasingly important role in the pathogenesis. To further investigate the role of the immune system, both cytokine gene polymorphisms and human leukocyte antigen (HLA) genotyping in patients with MTLE–HS were investigated.

**Methods:** The DNA samples of 100 patients with MTLE–HS and 201 healthy individuals were genotyped for cytokines (IL-6, IL-10, TNF- $\alpha$ , TGF- $\beta$ 1 and IFN- $\gamma$ ) and HLA using polymerase chain reaction (PCR)-SSP and SSO methods. The results were statistically analyzed in patient and healthy control groups and then according to the presence of febrile seizures (FS) in the patient group.

**Results:** Analysis of cytokine genotyping did not reveal any significant difference between patients with MTLE–HS and controls and patients with or without FS. However, the HLA DRB1\*13 allele was found to be more frequent in the patient population after Bonferroni correction.

**Conclusion:** This study suggests the possible role of HLA in the pathogenesis of MTLE–HS, although it failed to show any relationship with the cytokine system. However, data regarding the role of HLA are still lacking, and further studies are necessary to verify our results.

**Keywords:** Temporal lobe epilepsy, cytokine expression, HLA genotyping, hippocampal sclerosis

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### INTRODUCTION

Mesial temporal lobe epilepsy (MTLE) is a form of intractable epilepsy that may highly benefit from surgery in which hippocampal sclerosis (HS) is the most frequent pathological substrate. Histopathology of human epileptic hippocampi shows a specific neuronal loss in the areas of CA and gliosis (1). Formerly, the relationship between prolonged febrile seizures (FS) and the development of MTLE has been shown with regard to cytokine gene expression (2), although other studies have failed to replicate it (3,4,5). Because altered immunity due to the etio-pathogenesis of HS has been a point of interest, the role of proinflammatory cytokines and growth factors in the signaling process leading to a glial reaction have been investigated in animal models and human epilepsy (2,6,7,8). Despite recent studies, the pathogenesis of HS remains obscure, probably because of multifactorial reasons, including both environmental and genetic factors (9,10).

Previously, we performed HLA typing in patients with MTLE and found a significant difference in DR2, DR4, and DQ2 antigen distributions between the MTLE group and healthy population (11). Moreover, IL-1 $\beta$ / $\alpha$  gene polymorphism has been investigated in another group of patients with MTLE, but no significant difference has been observed (12).

To study the role of the immune system, we aimed to investigate other cytokine gene polymorphisms, including both pro- and anti-inflammatory cytokines (IL-6, IL-10, TNF- $\alpha$ , TGF- $\beta$ 1, IFN- $\gamma$ ), and HLA-DRB1 genotyping in patients with MTLE–HS.

### METHODS

In total, 100 consecutive patients with MTLE–HS who were admitted to the epilepsy outpatient clinic of Cerrahpaşa School of Medicine were studied. All these patients were surgical candidates because of medically intractable seizures and evaluated according to a standard protocol that included interictal and ictal video EEG recordings, cranial magnetic resonance imaging (MRI), and neuropsychological and psychiatric

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investigations. There were 68 female and 32 male patients, with a mean age of 25.8 years (range: 10–43) (Table 1). A history of FS was evident in 68% of the patients. HS was histopathologically confirmed in 70 patients who underwent anterior temporal lobectomy (the remaining were either under evaluation or in the waiting list for surgery). Informed consent was obtained from all the patients. The control group consisted of 201 healthy volunteers. This research was approved by the ethics committee of Istanbul University, Cerrahpaşa Faculty of Medicine (2006:18105).

DNA was extracted from the peripheral blood by using a standard method (13). Genotyping analysis of DRB1 loci and cytokine genes was performed at the Department of Medical Biology, Istanbul University. Cytokine genotypes were determined by the polymerase chain reaction (PCR)-SSP method using a commercial kit (One Lambda, Inc., Canoga Park, CA, USA) that contains sequence-specific primers for determining the following polymorphisms: TNF- $\alpha$  (–308 G/A), TGF- $\beta$ 1 (C/T codon 10, C/G codon 25), IL10 (–1082 G/A, –819 T/C, –592 A/C), IL-6 (–174 G/C), and IFN- $\delta$  (+874 A/T). DNA fragments corresponding to each cy-

tokine were amplified. The allele frequencies of specified cytokines were analyzed. In patients with MTLE, the correlations between the genotype of cytokines and the presence of FS were also investigated.

For analysis of HLA-DRB1, all the samples were typed at the DRB1 loci by PCR-based methods (PCR-SSO) at low resolution (14,15,16).

### Statistical Analysis

Statistical analysis was performed using Statistical Packages for the Social Sciences (SPSS Inc., Chicago, IL, USA) 12.0 version. Chi-square ( $\chi^2$ ) test with Bonferroni correction and binary logistic regression models were also utilized for analysis.

### RESULTS

Our results did not reveal any significant difference in the genotyping of pro- and anti-inflammatory cytokines between patients and controls. Further, allele distribution was not different in patients with or without FS. However, analyses of HLA-DRB1 showed a significant difference, where the DRB1\*13 allele was found to be more frequent in patients ( $p<0.05$ ), and it remained significant after Bonferroni correction.

### DISCUSSION

Although previous studies have suggested a role of genetic, immunological, and environmental factors (9,10,17), the etiopathogenesis of MTLE remains obscure. In this study, we aimed to enhance the understanding of the etiology and identify potential immune markers in the susceptibility of disease.

### Evaluation of Cytokines

Recent studies on epilepsy have suggested that the immune and inflammatory mechanisms play roles in some subgroups (18). In our study, the expressions of different cytokine genes (IL-6, IL-10, TNF- $\alpha$ , TGF- $\beta$ 1, and IFN- $\gamma$ ) and HLA-DR genotypes in Turkish patients with MTLE-HS were investigated to reveal the role of the immune system.

Cells in the central nervous system can produce cytokines upon activation (19), which may influence the transport systems by opening the blood–brain barrier (BBB). The administration of IL-1, IL-6, TNF- $\alpha$ , and IFN- $\gamma$  may also increase the endothelial permeability (20,21,22,23). Impaired BBB and inflammatory states are common features of neurological diseases associated with late-onset epilepsy (24). The involvement of inflammation in the pathogenesis of epilepsy and seizure-induced brain damage have been emphasized in many studies (17,18,25). Vezzani et al. (25) showed that the manipulation of pro- and anti-inflammatory cytokines can modify the outcome as well as the neuropathological consequences of experimentally induced seizures. Moreover, in patients with treatment-resistant epilepsy, increased levels of IL-6 and decreased levels of anti-inflammatory cytokine IL-1RA have been demonstrated (26). Recently, it has been shown that IL-1 $\beta$ , among others, is upregulated in pediatric patients with MTLE during the developmental phase of refractory seizures related to HS (27). The role of IL-1 $\beta$  in the pathogenesis of MTLE has been supported by the finding that the inflammatory effects of IL-1 $\beta$  lead to the upregulation of the PI3K/Akt/mTOR signaling pathway and could be suppressed by the administration of specific pathway inhibitors (28). The genetic susceptibility to inflammation has been found to be correlated with an increased risk of epilepsy (29). Weinberg et al. (30) explored the role of TNF- $\alpha$  and its two signaling receptors. They described a pro-ictogenic role of TNF- $\alpha$  mediated by TNFR1 and an anti-ictogenic role of this cytokine, probably mediated by TNFR2. The authors demonstrated that chronic induction of TNF- $\alpha$ , although inducing glial activation and BBB damage, did not provoke cell loss or seizures but enhanced the pathological consequences of seizures.

**Table 1.** Demographics and clinical characteristics of patients

Number of patients	100
Age (years)	25.8 (range:10–43)
Gender	68 females (68%) 32 male (32%)
Presence of febrile seizures	68 (68%)
Age at onset (years)	8.9 $\pm$ 5.8 (range: 1–28)
Disease duration (years)	18.4 $\pm$ 8.1 (range: 1–41)
Operation	70 patients (70%)
Age at operation (years)	25.4 (range:12–44)
Seizure characteristics	
Presence of GTCS	47 patients (47%)
Status epilepticus	13 patients (13%)
Presence of aura	65 patients (65%)
GTCS: generalized tonic-clonic seizures	

**Table 2.** HLA-DRB genotyping in patients and controls

	Patient n=200	Control n=402	$\chi^2$ test	p
DRB1*01	9	21	0.189	0.664
DRB1*03	26	30	3.770	0.052
DRB1*04	37	63	1.387	0.239
DRB1*07	21	35	0.376	0.540
DRB1*08	3	13	1.995	0.158
DRB1*09	0	7	fisher	0.100
DRB1*10	3	9	fisher	0.757
DRB1*11	40	87	0.307	0.579
DRB1*12	3	16	2.747	0.097
DRB1*13	22	27	5.199	0.037*
DRB1*14	17	29	0.169	0.681
DRB1*15	11	41	3.986	0.046
DRB1*16	8	24	1.091	0.296

n refers to the number of HLA alleles. \*p value was obtained from chi-square test and Bonferroni correction.

In many epilepsy models, acute seizures cause glial activation and increased expression of transcription factors and cytokines (31,32). The activated glia and elevated cytokines contribute to seizure-related hippocampal pathology such as neuronal death, reactive gliosis, and mossy fiber sprouting (33,34,35,36).

Differences in cytokine production are related to sequence variants in cytokine genes. Many of these polymorphisms occur in the regulatory regions, resulting in high or low cytokine production (37,38). We investigated different allelic variants in the promoter regions in pro-inflammatory (TNF- $\alpha$ , IFN- $\gamma$ , and IL6) and anti-inflammatory (IL10 and TGF $\beta$ 1) cytokine genes that have been identified previously (39). However, we did not find significant differences between patients with MTLE-HS and controls with regard to the genotype frequencies of IFN- $\gamma$ , TNF- $\alpha$ , TGF- $\beta$ 1, IL-6, and IL-10.

### Evaluation of HLA Genotype

HLA antigens and haplotypes have been described in connection with different epilepsies since the early 1970s; however, the results of such studies have been contradictory (40). We have reported a significantly high frequency of the HLA class II antigens DR4, DR7, and DQ2 in Turkish patients with MTLE-HS (11). Both our previous studies and several other studies have shown that HLA may play a role in different types of epilepsies (11,12,41). Moreover, the HLA-B\*1502 allele has been found to be strongly associated with carbamazepine-induced Stevens-Johnson syndrome and its related disease, toxic epidermal necrolysis, in Southeast Asian countries (42).

In this study, we investigated the polymorphic exons of the HLA-DR locus (exon 2 of the class II HLA locus) in patients with MTLE-HS based on our previous results, which suggested that DR4, DR7, and DQ2 are independent risk factors for MTLE-HS (11). In correlation with previous results, the present results revealed that the DRB1\*13 allele is more frequent in patients ( $p < 0.05$ ) than in healthy subjects (Table 2). Although the sample size is limited, our findings may suggest that a high frequency of HLA-DRB1\*13 allele plays a role in the induction of hyperexcitability of the epileptic tissue. Recently, the HLA-DRB1\*13:02 allele was found to be elevated in patients with MTLE-HS in comparison with controls, although the statistical significance was lost after Bonferroni correction (43). These findings may be taken into consideration as further evidence for immune-dependent genetic factors in the etiology of HS.

This study did not reveal any relationship between cytokine polymorphisms and human MTLE. However, the findings on HLA may suggest its role in the pathogenesis of HS, although this needs to be verified and requires further investigations, including a larger number of patients.

**Ethics Committee Approval:** Ethics committee approval was received for this study from the ethics committee of İstanbul University Cerrahpaşa School of Medicine.

**Informed Consent:** Written informed consent was obtained from patients who participated in this study.

**Peer-review:** Externally peer-reviewed.

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F.S.O.; Writing Manuscript – A.A., Ç.Ö., F.S.O.; Critical Review – A.A., Ç.Ö., F.S.O.; Other – A.A., Ç.Ö., M.U., F.S.O.

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### REFERENCES

- Özkara Ç, Aronica E. Hippocampal sclerosis. *Handb Clin Neurol* 2012; 108:621-639. [CrossRef]
- Kanemoto K, Kawasaki J, Miyamoto T, Obayashi H, Nishimura M. Interleukin (IL)1beta, IL-1alpha, and IL-1 receptor antagonist gene polymorphisms in patients with temporal lobe epilepsy. *Ann Neurol* 2000; 47:571-574.
- Tilgen N, Pfeiffer H, Cobilanschi J, Rau B, Horvath S, Elger CE, Propping P, Heils A. Association analysis between the human interleukin 1beta (-511) gene polymorphism and susceptibility to febrile convulsions. *Neurosci Lett* 2002; 334:68-70. [CrossRef]
- Tsai FJ, Hsieh YY, Chang CC, Lin CC, Tsai CH. Polymorphisms for interleukin 1b exon 5 and interleukin 1 receptor antagonist in Taiwanese Children with febrile convulsions. *Arch Pediatr Adolesc Med* 2002; 156:545-548. [CrossRef]
- Kira R, Torisu H, Takemoto M, Nomura A, Sakai Y, Sanefuji M, Sakamoto K, Matsumoto S, Gondo K, Hara T. Genetic susceptibility to simple febrile seizures: Interleukin-1b promoter polymorphisms are associated with sporadic cases. *Neurosci Lett* 2005; 384:239-244. [CrossRef]
- Straussberg R, Amir J, Harel L, Punsky I, Bessler H. Pro and antiinflammatory cytokines in children with febrile convulsions. *Pediatr Neurol* 2001; 24:49-53. [CrossRef]
- Peltola J, Laaksonen J, Haapala AM, Hurme M, Rainesalo S, Keränen T. Indicators of inflammation after recent tonic-clonic epileptic seizures correlate with plasma interleukin-6 levels. *Seizure* 2002; 11:44-46. [CrossRef]
- Goldstein KR, Bhatt R, Barton BE, Zalcman SS, Rameshwar P, Siegel A. Effects of hemispheric lateralization and site specificity on immune alterations induced by kindled temporal lobe seizures. *Brain Behav Immun* 2002; 16:706-719. [CrossRef]
- Berkovic SF, Mulley JC, Scheffer IE, Petrou S. Human epilepsies: interaction of genetic and acquired factors. *Trends Neurosci* 2006; 29:391-397. [CrossRef]
- Tan NC, Mulley JC, Scheffer IE. Genetic dissection of the common epilepsies. *Curr Opin Neurol* 2006; 19:157-163. [CrossRef]
- Özkara C, Altıntaş A, Yılmaz E, Eskazan E, Erkol G, Ozyurt E, Erdogan E, Kuday C. An association between mesial temporal lobe epilepsy with hippocampal sclerosis and human leucocyte antigens. *Epilepsia* 2002; 43:236-239. [CrossRef]
- Ozkara C, Uzan M, Tanriverdi T, Baykara O, Ekinci B, Yeni N, Kafadar A, Buyru N. Lack of Association between IL-1  $\beta/\alpha$  Gene Polymorphisms and Temporal Lobe Epilepsy with Hippocampal Sclerosis. *Seizure* 2006; 15:288-291. [CrossRef]
- Gustincich S, Manfioletti G, Del Sal G, Schneider C, Carninci P. A fast method for high quality genomic DNA extraction from whole human blood. *Bio Techniques* 1991; 11:298-302.
- Buyse I, Decorte R, Baens M, Cuppens H, Semana G, Emonds MP, Marynen P, Cassiman JJ. Rapid DNA typing of class II antigens using the PCR and reverse dot blot hybridization. *Tissue Antigens* 1993; 41:1-14. [CrossRef]
- Anholts JD, Verduyn W, Parlevliet A, Doxiadis II, D'Amaro J, Giphart MJ, Persijn GG, Schreuder GM. Irregular PCR-SSO hybridization patterns reveal seven new HLA-DRB1 alleles related to DR2, DR3, DR6, DR8 and DR11; Implications for sequence specific priming. *Hum Immunol* 1995; 42:15-22. [CrossRef]
- Jordan F, McWhinnie AJ, Turner S, Gavira N, Calvert AA, Cleaver SA, Holman RH, Goldman JM, Madrigal JA. Comparison of HLA-DRB1 typing by DNA-RFLP, PCR-SSO and PCR-SSP methods and their application in providing matched unrelated donors for bone marrow transplantation. *Tissue Antigens* 1995; 45:103-110. [CrossRef]
- Vezzani A, Balosso S, Ravizza T. The role of cytokines in the pathophysiology of epilepsy. *Brain Behav Immun* 2008; 22:797-803. [CrossRef]
- Vezzani A, French J, Bartfai T, Baram TZ. The role of inflammation in epilepsy. *Nat Rev Neurol* 2011; 7:31-40. [CrossRef]

19. de Boer AG, Breimer DD. Cytokines and blood-brain barrier permeability. *Prog Brain Res* 1998; 115:425-451. [\[CrossRef\]](#)
20. Schilling L, Wahl M. Mediators of cerebral edema. *Adv Exp Med Biol* 1999; 474:123-141. [\[CrossRef\]](#)
21. Laflamme N, Lacroix S, Rivest S. An essential role of interleukin-1 $\beta$  in mediating NF- $\kappa$ B activity and COX-2 transcription in cells of the blood-brain barrier in response to a systemic and localized inflammation but not during endotoxemia. *J Neurosci* 1999; 19:10923-10930.
22. Freyer D, Manz R, Ziegenhorn A, Weih M, Angstwurm K, Döcke WD, Meisel A, Schumann RR, Schönfelder G, Dirnagl U, Weber JR. Cerebral endothelial cells release TNF- $\alpha$  after stimulation with cell walls of *Streptococcus pneumoniae* and regulate inducible nitric oxide synthase and ICAM-1 expression via autocrine loops. *J Immunol* 1999; 163:4308-4314.
23. Rivest S. What is the cellular source of prostaglandins in the brain in response to systemic inflammation? Facts and controversies. *Mol Psychiatry* 1999; 4:500-507. [\[CrossRef\]](#)
24. Krizanac-Bengez L, Mayberg MR, Janigro D. The cerebral vasculature as a therapeutic target for neurological disorders and the role of shear stress in vascular homeostasis and pathophysiology. *Neurol Res* 2004; 26:846-853. [\[CrossRef\]](#)
25. Vezzani A, Moneta D, Richichi C, Aliprandi M, Burrows SJ, Ravizza T, Perego C, De Simoni MG. Functional role of inflammatory cytokines and antiinflammatory molecules in seizures and epileptogenesis. *Epilepsia* 2002; 5:30-35. [\[CrossRef\]](#)
26. Hulkkonen J, Koskikallio E, Rainesalo S, Keränen T, Hurme M, Peltola J. The balance of inhibitory and excitatory cytokines is differently regulated in vivo and in vitro among therapy resistant epilepsy patients. *Epilepsy Res* 2004; 59:199-205. [\[CrossRef\]](#)
27. Omran A, Peng J, Zhang C, Xiang QL, Xue J, Gan N, Kong H, Yin F. Interleukin-1 $\beta$  and microRNA-146a in an immature rat model and children with mesial temporal lobe epilepsy. *Epilepsia* 2012; 53:1215-1224. [\[CrossRef\]](#)
28. Xiao Z, Peng J, Yang L, Kong H, Yin F. Interleukin-1 $\beta$  plays a role in the pathogenesis of mesial temporal lobe epilepsy through the PI3K/Akt/mTOR signaling pathway in hippocampal neurons. *J Neuroimmunol* 2015; 282:110-117. [\[CrossRef\]](#)
29. Choi J, Koh S. Role of brain inflammation in epileptogenesis. *Yonsei Med J* 2008; 49:1-18. [\[CrossRef\]](#)
30. Weinberg MS, Blake BL, McCown TJ. Opposing actions of hippocampus TNF $\alpha$  receptors on limbic seizure susceptibility. *Exp Neurol* 2013; 247:429-437. [\[CrossRef\]](#)
31. Plata-Salamán CR, Ilyin SE, Turrin NP, Gayle D, Flynn MC, Romanovitch AE. Kindling modulates the IL-1 $\beta$  system, TNF- $\alpha$ , TGF- $\beta$ 1, and neuropeptide mRNAs in specific brain regions. *Brain Res Mol Brain Res* 2000; 75:248-258. [\[CrossRef\]](#)
32. Zimmer LA, Ennis M, Shipley MT. Soman-induced seizures rapidly activate astrocytes and microglia in discrete brain regions. *J Comp Neurol* 1997; 378:482-492.
33. McNamara JO. Cellular and molecular basis of epilepsy. *J Neurosci* 1994; 14:3413-3425.
34. Parent JM, Yu TW, Leibowitz RT, Geschwind DH, Sloviter RS, Lowenstein DH. Dentate granule cell neurogenesis is increased by seizures and contributes to aberrant network reorganization in the adult rat hippocampus. *J Neurosci* 1997; 17:3727-3738.
35. Jankowsky JL, Patterson PH. The role of cytokines and growth factors in seizures and their sequelae. *Prog Neurobiol* 2001; 63:125-149. [\[CrossRef\]](#)
36. Koh S, Storey TW, Santos TC, Mian AY, Cole AJ. Early-life seizures in rats increase susceptibility to seizure-induced brain injury in adulthood. *Neurology* 2001; 57:22-28.
37. Bidwell J, Keen L, Gallagher G, Kimberly R, Huizinga T, McDermott MF, Oksenberg J, McNicholl J, Pociot F, Hardt C, D'Alfonso S. Cytokine gene polymorphism in human disease: on-line databases. *Genes Immun* 1999; 1:3-19. [\[CrossRef\]](#)
38. Bidwell J, Keen L, Gallagher G, Kimberly R, Huizinga T, McDermott MF, Oksenberg J, McNicholl J, Pociot F, Hardt C, D'Alfonso S. Cytokine gene polymorphism in human disease: on-line databases, Supplement 1. *Genes Immun* 2001; 2:61-70. [\[CrossRef\]](#)
39. Wilson AG, Symons JA, McDowell TL, McDevitt HO, Duff GW. Effects of a polymorphism in the human tumor necrosis factor  $\alpha$  promoter on transcriptional activation. *Proc Natl Acad Sci U S A* 1997; 94:3195-3199. [\[CrossRef\]](#)
40. Eeg-Olofsson O. Virological and immunological aspects of seizure disorders. *Brain Dev* 2003; 25:9-13. [\[CrossRef\]](#)
41. Durner M, Janz D, Zingsem J, Greenberg DA. Possible association of JME with HLA-DRW6. *Epilepsia* 1992; 33:814-816. [\[CrossRef\]](#)
42. Chung WH, Hung SI, Hong HS, Hsih MS, Yang LC, Ho HC, Wu JY, Chen YT. Medical genetics: a marker for Stevens-Johnson syndrome. *Nature* 2004; 428:486. [\[CrossRef\]](#)
43. Horta WG, Paradela E, Figueiredo A, Meira ID, Pereira VC, Rego CC, Oliveira R, Andraus ME, de Lacerda GC, Moura P, de Souza JP, Paiva CL, Alves-Leon SV. Genetic association study of the HLA class II alleles DRB1, DQA1, and DQB1 in patients with pharmacoresistant temporal lobe epilepsy associated with mesial hippocampal sclerosis. *Seizure* 2015; 31:7-11. [\[CrossRef\]](#)